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Glibenclamide, ATP and Metformin Increases the Expression of Human Bile Salt Export Pump ABCB11

ABSTRACT

Background:

Bile Salt Export Pump (BSEP/ABCB11) is important in the maintenance of the enterohepatic circulation of bile acids and drugs. Drugs such as rifampicin, glibenclamide inhibit BSEP. Progressive Familial Intrahepatic Cholestasis Type-2, a lethal pediatric disease, some forms of intrahepatic cholestasis of pregnancy, and drug-induced cholestasis are associated with BSEP dysfunction.

Methods:

We started with a bioinformatic approach to identify the relationship between ABCB11 and other proteins, microRNAs, and drugs. Microarray data set of the liver samples from ABCB11 knockout mice was analyzed by GEO2R tool. Differentially expressed gene pathway enrichment analysis was done by ClueGo v2.5.5 app from Cytoscape. Protein-protein interaction network was constructed by STRING application in Cytoscape. Networks were analyzed using the Cytoscape software v3.7.1. CyTargetLinker v4.1.0 was used to screen the transcription factors, microRNAs and drugs. Predicted drugs were validated on human liver cell line, HepG2. BSEP expression was quantified by Real Time PCR and Western Blot.

Results:

ABCB11 knockout in mice was associated with a predominant upregulation and downregulation of genes associated with cellular component movement and sterol metabolism respectively. We further identified the hub genes in the network. Genes related to immune activity, cell signaling and fatty acid metabolism were dysregulated. We further identified drugs (glibenclamide and ATP) and a total of 14 microRNAs targeting the gene. Western Blot and Real Time PCR analysis confirmed the upregulation of BSEP on the treatment of HepG2 cells with glibenclamide, ATP, and metformin.

Conclusion:

The differential expression of cell signaling genes and those related to immune activity in ABCB11 KO animals may be secondary to cell injury. We have found glibenclamide, ATP, and metformin upregulates BSEP. The mechanisms involved and the clinical relevance of these findings need to be investigated.

Keywords:

BSEP/ABCB11, ABCB11-KO, Insilico, upregulation, HepG2, glibenclamide, ATP, and metformin, Nuclear Receptors

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INTRODUCTION

The Bile Salt Export Pump (BSEP), the major bile salt transporter in the liver canalicular membrane, is coded by *ABCB11* gene, and mutations in this gene cause Progressive Familial Intrahepatic Cholestasis Type- 2 (PFIC-2) (1,2). Besides PFIC-2, mutations or insufficiency of BSEP is associated with a variety of diseases such as drug-induced cholestasis, pregnancy induced cholestasis, cryptogenic cholestasis, cholangiocarcinoma and hepatocellular carcinoma, which are cancers of the liver(3–7). Naturally, ABCB11 expression is induced by bile salts and is mediated by FXR- RXR heterodimer (8). Here in this pilot study we explored in-silico the interactions/networks around ABCB11. We wanted to identify the genes, drugs, microRNAs which might influence the expression of ABCB11. Drugs which could upregulate ABCB11 expression may be useful in ABCB11 haploinsufficiency and inhibition of the pump could result in the accumulation of toxic bile salts inside hepatocytes. Modulation of ABCB11 expression could be clinically beneficial in a variety of medical conditions.

MATERIALS & METHODS

Identification of differentially expressed genes: We analyzed the microarray data set of the liver samples from ABCB11 knockout mice (GSE 70179) using GEO2R online tool from NCBI (9). All differentially expressed genes (DEGs) were filtered with two criteria: -1> $\log_2$FC >+1 and adj. p-value <0.05.

Pathway enrichment analysis: To identify DEGs which are significant, pathway enrichment analysis was conducted using the ClueGo v2.5.5 app from Cytoscape(10). ClueGo constructed and compared networks of functionally related GO terms with kappa statistics, which was adjusted at >0.4 in this study.

Identification of hub genes and subnetwork analysis: The protein-protein interaction (PPI) networks were built by the Search Tool for the Retrieval of Interacting Genes (STRING v11.0) (11)and Cytoscape v3.7.1 software. The Molecular Complex Detection (MCODE), app from Cytoscape was used to screen modules of the PPI network with degree cut-off = 2, node score cut-off = 0.2, k-core = 2, and maximum depth = 100. The hub genes were identified by the CytoHubba app. The top 10 nodes were considered as notable hub genes and displayed.

Identification of transcription factor and drug target: CyTargetLinker app from Cytoscape was used to identify the transcription factor (TF) and drugs targeting the dysregulated genes of the selected category. We drew Homo sapiens TF-target interactions linkset from database (ENCODE) (12) and drug-target interactions linkset from the database (DrugBank)(13). The networks were visualized and analyzed using the Cytoscape software v3.7.1

Cell Culture: HepG2 cells were grown in high glucose DMEM (Hi-Media Lab, Mumbai, Cat. # AL111-500ML) supplemented with 10% fetal bovine serum (CellClone, Genetix Biotech Asia, New Delhi, Cat.# CCS-500-SA-U), penicillin and streptomycin (Hi-Media, Mumbai Cat. # A018-5X100ML). When cells became 80% confluent, they were treated with glibenclamide (500ng/mL) (14), metformin (25 mg/L) (15) and ATP (1mM) for 48h. Cells were harvested after 48h cells were harvested for total protein and RNA.

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Western Blot: Total proteins from HepG2 cells were prepared and run on 10% SDS-PAGE and transferred to a PVDF membrane using a transfer apparatus following the standard protocols (Bio-Rad). After incubation with 5% nonfat milk in TBST (10 mM Tris, pH 8.0, 150 mM NaCl, 0.5% Tween 20) for 1h the membrane was washed once with TBST and incubated with antibodies against human ABCB11 (Affinity, Catalog #DF 9278) 1: 2000 dilution; human β-actin (Santa Cruz Cat.# SC4778), dilution 1:1000. The membrane was washed and incubated with a 1:5000 dilution of horseradish peroxidase-conjugated anti-rabbit (Santa Cruz Cat# SC-2004)/anti-mouse antibodies (Cat.#SC-2005) for 2 h. Blots were washed with TBST four times and developed with the ECL system (Bio-Rad, US Cat.#170-5060) according to the 'manufacturer's protocol.

Real Time PCR: Total RNA was isolated using NucleoZOL (Takara Cat. No. 740404.200) following manufacturer's instruction. cDNA was prepared from (deoxyribonuclease treated) total RNA using RevertAid Reverse Transcriptase (Thermo Cat. No. EP0441) following the 'manufacturer's instructions. Real Time PCR was done with unique oligonucleotide primers targeting ABCB11 gene using GoTaq® qPCR Master Mix (Promega Cat. No. A6001) following 'manufacturer's instructions on a Veriti Thermo Cycler from Applied Biosystems Waltham, Massachusetts, USA and data was acquired using the software platform associated with the same machine.

RESULTS

Expression of 560 genes changed significantly following ABCB11-KO in 1.5 m mice

Gene expression profile ABCB11 knockdown dataset GSE 70179 from GEO datasets were analysed with GEO2R tool. Genes with >2-fold change in expression value and <0.05 adjusted p value was filtered. Identified differentially expressed genes (DEG) from the GSE dataset were classified in two groups - upregulated (375 genes) and downregulated (185 genes) (Supp.Table-1). Gene ontology analysis was performed for functional analysis of DEGs by using ClueGo app from Cytoscape. PPIs of DEGs were constructed using STRING database showed an upregulation of genes related to cellular transport (pink colored nodes), and these nodes were also shared by Toll Like Receptor (TLR) signalling (Fig-1). Downregulated genes were involved in metabolic pathways (sterol, carbohydrate, alcohol, etc) (Supp. Table-2). We next identified top hub genes in PPI network using CytoHubba app from Cytoscape (Table-1). Immunologically important genes were among the top ranked upregulated hub genes (Fig-2a) downregulated group majorly represents cell signaling and fatty acid metabolism (Fig-2b). Epidermal Growth Factor Receptor (EGFR) ranked first among the genes involved in signaling pathways. Kinases play a role in the transcription, activity, or intracellular localization of ABC transporters as do protein interactions(16). Proteins which are interacting with ABCB11 are represented in Supp. Fig-1 which includes nuclear receptors NR1H4 and NR0B2. Most proteins were associated with bile acid metabolism and transport.

Sub-network analysis was performed using the Molecular Complex Detection (MCODE) app from Cytoscape (Fig-3), and CMPK2, ACTG1, and SSTR2 emerged as seed nodes among upregulated (Table -2). Among downregulated gene groups only one subnetwork was found to be significant which had three genes *MIA3* (which codes a protein which is important in the transport of cargos that are too large to fit into COPII-coated vesicles such as collagen VII), IGFBP4 (a protein that binds to both insulin-like growth factors and modifies their functions) and NOTUM (a carboxylesterase that acts as a key negative regulator of the Wnt signaling pathway by specifically mediating depalmitoleoylation of WNT proteins).

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Using CyTargetLinker app from Cytoscape identified two drugs, glibenclamide, and ATP, directly targeting ABCB11. We subsequently looked for microRNAs [Target-scan database](17) that were associated with ABCB11, and a total of 14 microRNAs were identified targeting the gene (Fig-4). Transcription factors and microRNAs targeting ABCB11 and interacting partners are represented in Supp. Fig-2.

Glibenclamide ATP and Metformin upregulates ABCB11

We evaluated in-vitro, the effect of three drugs, two of which were bioinformatically predicted (Glibenclamide, ATP) and one based on literature (18). We found all the three compounds upregulating ABCB11 expression based on Real Time PCR, and this was confirmed by Western Blot (Fig-5).

DISCUSSION

We identified several immunologically important genes being upregulated ABCB11 deficiency. The reason could be liver cell injury secondary to bile salt accumulation, which triggers sterile immune response(19,20) and the downregulation of transport protein and metabolically important genes could be because of decreased liver function following damage. A regenerative response follows cell injury, and a host of genes involved in regeneration are upregulated (21–23) however, it appears that bile salts in the absence of BSEP hamper the regenerative response reflected by dysregulated collagen transporting protein MIA3 and NOTUM a protein involved in Wnt signaling. it's also possible that EGFR is dysregulated via accumulating bile salts mediated by STAT3(24). We have observed an upregulation of ABCB11 in liver cell line (HepG2) on treatment with glibenclamide, metformin, and ATP. Its upregulation may be a compensatory mechanism in the case of glibenclamide and metformin because these drugs are known to inhibit ABCB11(25). Metformin is known to interfere with ABCB11 function mediated through AMPK-FXR crosstalk(18) involving metformin induced FXR phosphorylation. ATP acts through ATP receptors on hepatocytes(26,27). ATP is known to cross plasma membrane(28) and this can act via AMPK. However, ATP has a very short half-life (29), and it may be converted to ADP, which can activate AMPK(30). In a recent report, metformin was shown to suppress ABCB11 expression, which is not in agreement with our observation, however, they performed their experiment on primary human hepatocytes, and they have also treated their cells with DMSO (31). However, we are still unable to explain the observed difference. In conclusion, we need more experiments to determine the mechanisms of action of these drugs on the upregulation of ABCB11. Many changes in gene expression following ABCB11 could be secondary to stress, immune and regenerative responses following hepatocyte injury in mice liver.

BIBLIOGRAPHY

1. Strautnieks SS, Byrne JA, Pawlikowska L, Cebecauerová D, Rayner A, Dutton L, et al. Severe bile salt export pump deficiency: 82 different ABCB11 mutations in 109 families. Gastroenterology. 2008 Apr;134(4):1203–1214.

2. Childs S, Yeh RL, Georges E, Ling V. Identification of a sister gene to P-glycoprotein. Cancer Res. 1995 May 15;55(10):2029–2034.

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3. Davit-Spraul A, Gonzales E, Jacquemin E. NR1H4 analysis in patients with progressive familial intrahepatic cholestasis, drug-induced cholestasis or intrahepatic cholestasis of pregnancy unrelated to ATP8B1, ABCB11 and ABCB4 mutations. Clin Res Hepatol Gastroenterol. 2012 Dec;36(6):569–573.

4. Dixon PH, van Mil SWC, Chambers J, Strautnieks S, Thompson RJ, Lammert F, et al. Contribution of variant alleles of ABCB11 to susceptibility to intrahepatic cholestasis of pregnancy. Gut. 2009 Apr;58(4):537–544.

5. Scheimann AO, Strautnieks SS, Knisely AS, Byrne JA, Thompson RJ, Finegold MJ. Mutations in bile salt export pump (ABCB11) in two children with progressive familial intrahepatic cholestasis and cholangiocarcinoma. J Pediatr. 2007 May;150(5):556–559.

6. Knisely AS, Strautnieks SS, Meier Y, Stieger B, Byrne JA, Portmann BC, et al. Hepatocellular carcinoma in ten children under five years of age with bile salt export pump deficiency. Hepatology. 2006 Aug;44(2):478–486.

7. Vitale G, Gitto S, Raimondi F, Mattiaccio A, Mantovani V, Vukotic R, et al. Cryptogenic cholestasis in young and adults: ATP8B1, ABCB11, ABCB4, and TJP2 gene variants analysis by high-throughput sequencing. J Gastroenterol. 2018 Aug;53(8):945–958.

8. Plass JRM, Mol O, Heegsma J, Geuken M, Faber KN, Jansen PLM, et al. Farnesoid X receptor and bile salts are involved in transcriptional regulation of the gene encoding the human bile salt export pump. Hepatology. 2002 Mar;35(3):589–596.

9. GEO2R - GEO - NCBI [Internet]. [cited 2020 Jul 24]. Available from: https://www.ncbi.nlm.nih.gov/geo/geo2r/?acc=GSE%2070179

10. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003 Nov;13(11):2498–2504.

11. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019 Jan 8;47(D1):D607–D613.

12. ENCODE Project Consortium. The ENCODE (encyclopedia of DNA elements) project. Science. 2004 Oct 22;306(5696):636–640.

13. Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. Nucleic Acids Res. 2018 Jan 4;46(D1):D1074–D1082.

14. Matsuda A, Kuzuya T, Sugita Y. Plasma levels of glibenclamide in diabetic patients during its routine clinical administration determined by a specific radioimmunoassay. … and metabolic research. 1983;

15. Kajbaf F, De Broe ME, Lalau J-D. Therapeutic concentrations of metformin: A systematic review. Clin Pharmacokinet. 2016 Apr;55(4):439–459.

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16. Crawford RR, Potukuchi PK, Schuetz EG, Schuetz JD. Beyond Competitive Inhibition: Regulation of ABC Transporters by Kinases and Protein-Protein Interactions as Potential Mechanisms of Drug-Drug Interactions. Drug Metab Dispos. 2018 Mar 7;46(5):567–580.

17. Agarwal V, Bell GW, Nam J-W, Bartel DP. Predicting effective microRNA target sites in mammalian mRNAs. Elife. 2015 Aug 12;4.

18. Lien F, Berthier A, Bouchaert E, Gheeraert C, Alexandre J, Porez G, et al. Metformin interferes with bile acid homeostasis through AMPK-FXR crosstalk. J Clin Invest. 2014 Mar;124(3):1037–1051.

19. Allen K, Jaeschke H, Copple BL. Bile acids induce inflammatory genes in hepatocytes: a novel mechanism of inflammation during obstructive cholestasis. Am J Pathol. 2011 Jan;178(1):175–186.

20. Woolbright BL, Jaeschke H. The impact of sterile inflammation in acute liver injury. J Clin Transl Res. 2017 Feb 12;3(Suppl 1):170–188.

21. Dransfeld O, Gehrmann T, Köhrer K, Kircheis G, Holneicher C, Häussinger D, et al. Oligonucleotide microarray analysis of differential transporter regulation in the regenerating rat liver. Liver Int. 2005 Dec 1;25(6):1243–1258.

22. White P, Brestelli JE, Kaestner KH, Greenbaum LE. Identification of transcriptional networks during liver regeneration. J Biol Chem. 2005 Feb 4;280(5):3715–3722.

23. Li S-C, Wang F-S, Yang Y-L, Tiao M-M, Chuang J-H, Huang Y-H. Microarray Study of Pathway Analysis Expression Profile Associated with MicroRNA-29a with Regard to Murine Cholestatic Liver Injuries. Int J Mol Sci. 2016 Mar 1;17(3).

24. Mair M, Zollner G, Schneller D, Musteanu M, Fickert P. Signal transducer and activator of transcription 3 protects from liver injury and fibrosis in a mouse model of sclerosing cholangitis. Gastroenterology. 2010;

25. Krivoy N, Zaher A, Yaacov B, Alroy G. Fatal toxic intrahepatic cholestasis secondary to glibenclamide. Diabetes care. 1996;

26. Schlosser SF, Burgstahler AD, Nathanson MH. Isolated rat hepatocytes can signal to other hepatocytes and bile duct cells by release of nucleotides. Proc Natl Acad Sci USA. 1996 Sep 3;93(18):9948–9953.

27. Charest R, Blackmore PF, Exton JH. Characterization of responses of isolated rat hepatocytes to ATP and ADP. J Biol Chem. 1985 Dec 15;260(29):15789–15794.

28. Chaudry IH. Does ATP cross the cell plasma membrane. Yale J Biol Med. 1982 Feb;55(1):1–10.

29. Skrabanja ATP, Bouman EAC, Dagnelie PC. Potential value of adenosine '5'-triphosphate (ATP) and adenosine in anaesthesia and intensive care medicine. Br J Anaesth. 2005 May 1;94(5):556–562.

30. Hardie DG. AMP-activated protein kinase: an energy sensor that regulates all aspects of cell function. Genes Dev. 2011 Sep 15;25(18):1895–1908.

31. Garzel B, Hu T, Li L, Lu Y, Heyward S, Polli J, et al. Metformin disrupts bile acid efflux by repressing bile salt export pump expression. Pharm Res. 2020 Jan 6;37(2):26.

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Disclosures:

Conflict of interest: None of the authors listed in this publication have any conflict of interest financial or otherwise, pertinent to the work, concepts, results or ideas described in this manuscript which is being submitted for favor of publication. Sources of funding which supported this work (SERB and DBT, Government of India) are mentioned below and they are governmental research funding agencies who do not have any conflict/conflicts of interests with reference to the content of this manuscript.

Ethical approval and Informed consent: No human patient or animal material/specimens were used in this work which requires an Institutional Ethics Committee/Institutional Review Board approval to the best of our knowledge.

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Figure-1

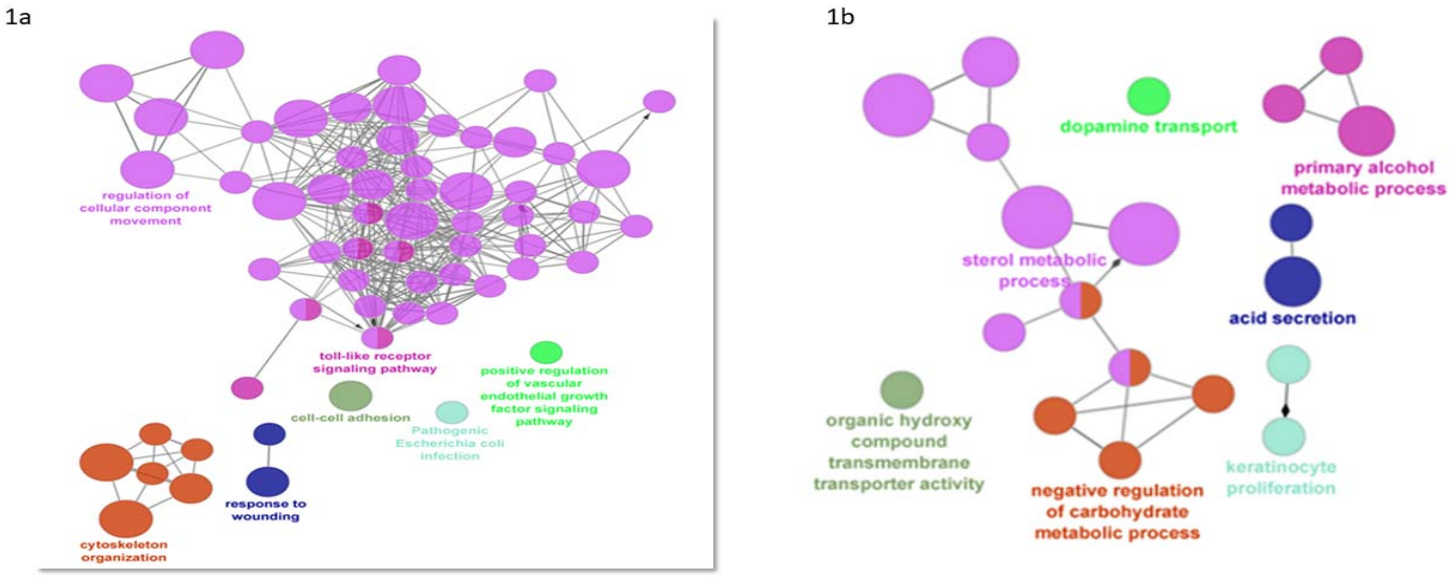


Figure-1

Protein-protein interaction networks (PPIs) of DEGs were constructed using STRING database. Gene ontology analysis was performed for functional analysis of DEGs by using ClueGo app from Cytoscape. This app allows simultaneous analysis of multiple annotation and ontology sources. Functionally grouped network is represented Figure 1a (upregulated genes) 1b (downregulated genes) . The node size represents enrichment significance and connections are based on kappa score (> 0.4). In upregulated gene group maximum number of nodes which are in pink color represent the cellular component movement. These nodes are shared by toll like receptor signaling pathway.

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Figure-2

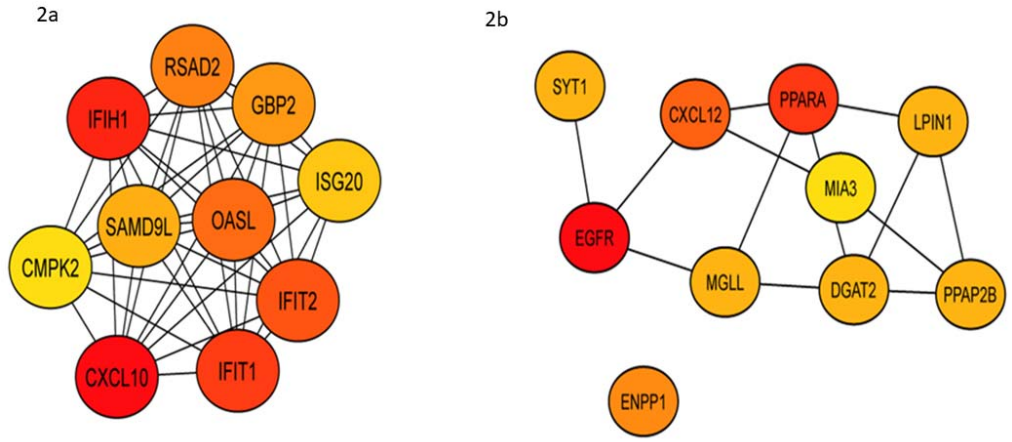


Figure-2

We identified top hub genes in PPI network of both upregulated and downregulated genes by using CytoHubba app from Cytoscape. We observed that the top ranked hub genes in the upregulated group were mainly related to immune activity (2a). The top hub genes in downregulated group were associated with cell signaling and fatty acid metabolism. The EGFR emerged as the top hub gene, a growth factor receptor which is crucial factor several cell signaling pathways (2b).

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Figure-3

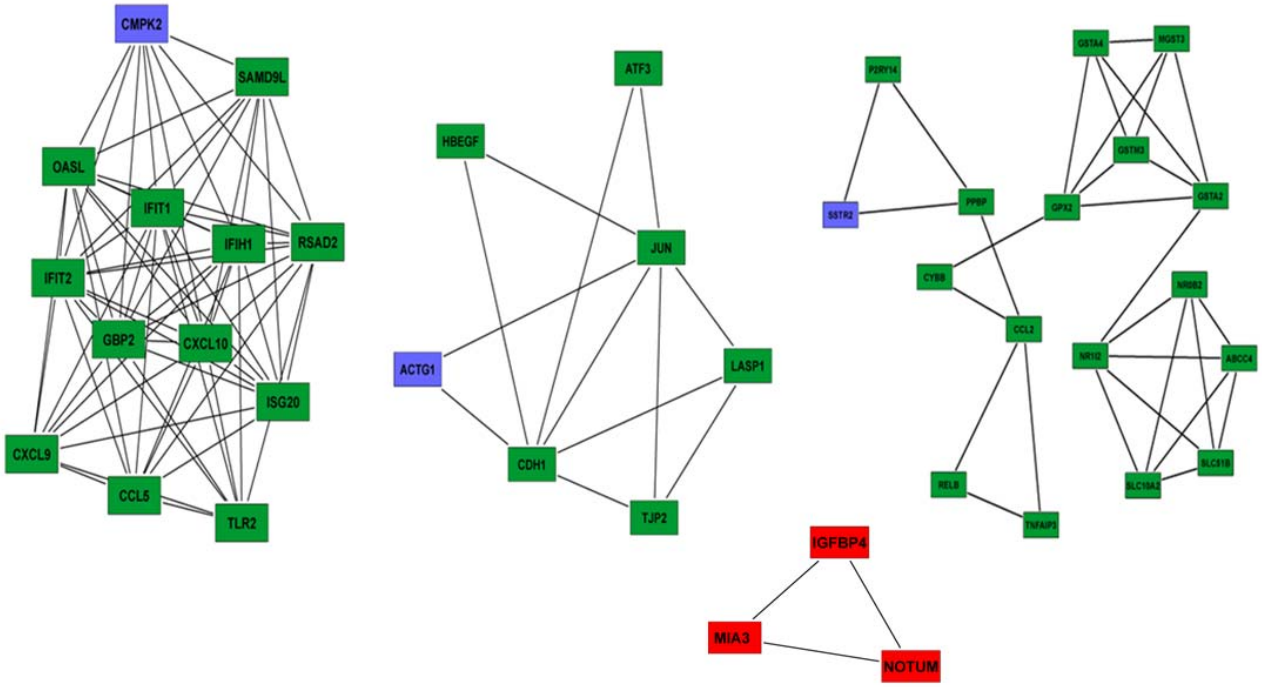


Figure-3

Sub-network analysis was conducted by using the Molecular Complex Detection (MCODE) app from Cytoscape. Top sub-networks on the basis of MCODE score (Degree cut-off= 2, node score cut-off = 0.2, k-core = 2 and max. depth = 100). Upregulated gene group clusters, we identified seed nodes (CMPK2, ACTG1 and SSTR2) in the network (green and blue). In downregulated gene group, we identified only one subnetwork which qualified cut off criteria. Three genes in this sub-network was identified: MIA3, IGFBP4 and NOTUM (red).

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Figure-4

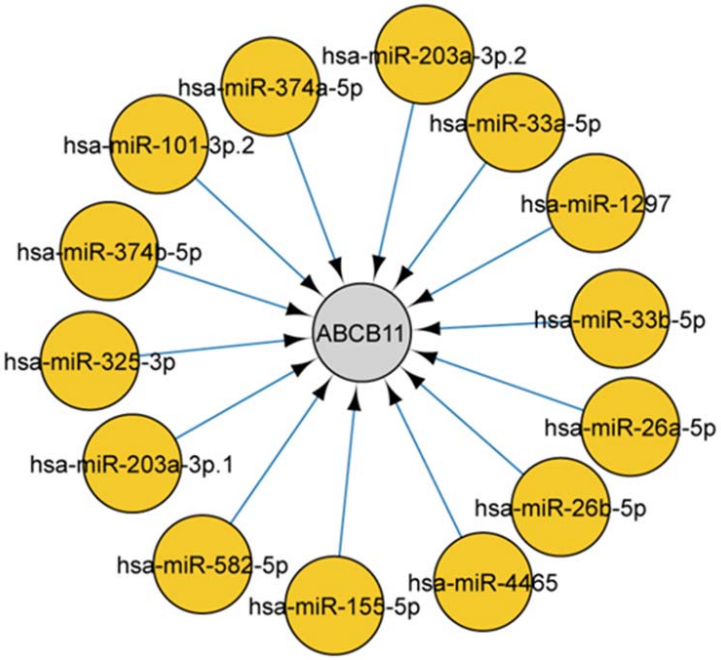


Figure-4

Cytoscape app CyTargetlinker version 4.1.0[6] was used to screen microRNAs using the Target-scan database. We identified microRNAs that were associated with ABCB11. In total 14 microRNA identified targeting the gene.

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Figure-5

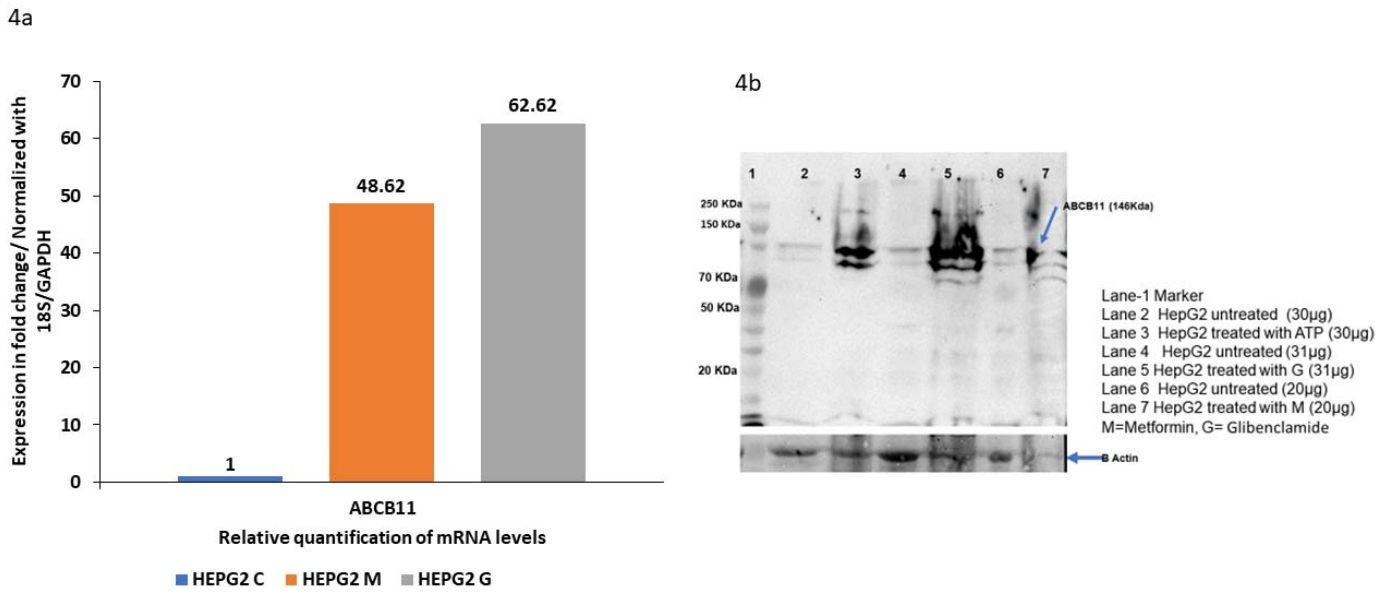
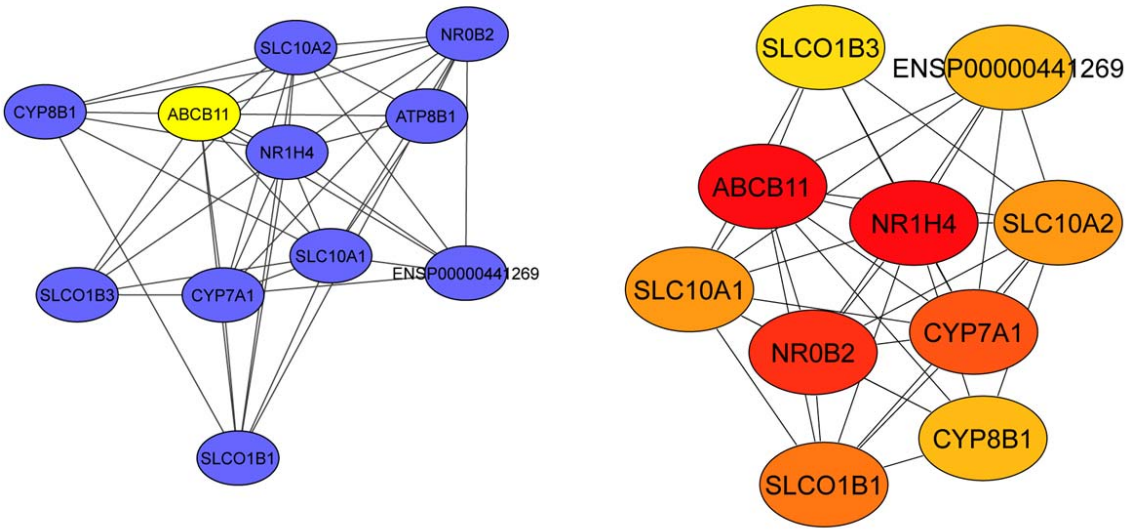


Figure-5

Three drugs, two of which were bioinformatically predicted (Glibenclamide, ATP) and one (Metformin) based on evidence from literature. All the three compounds upregulated ABCB11 expression based on Real Time PCR data. This was further confirmed by Western Blot against anti-human ABCB11 antibody as the primary antibody.

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Supp-Fig-1 a,b

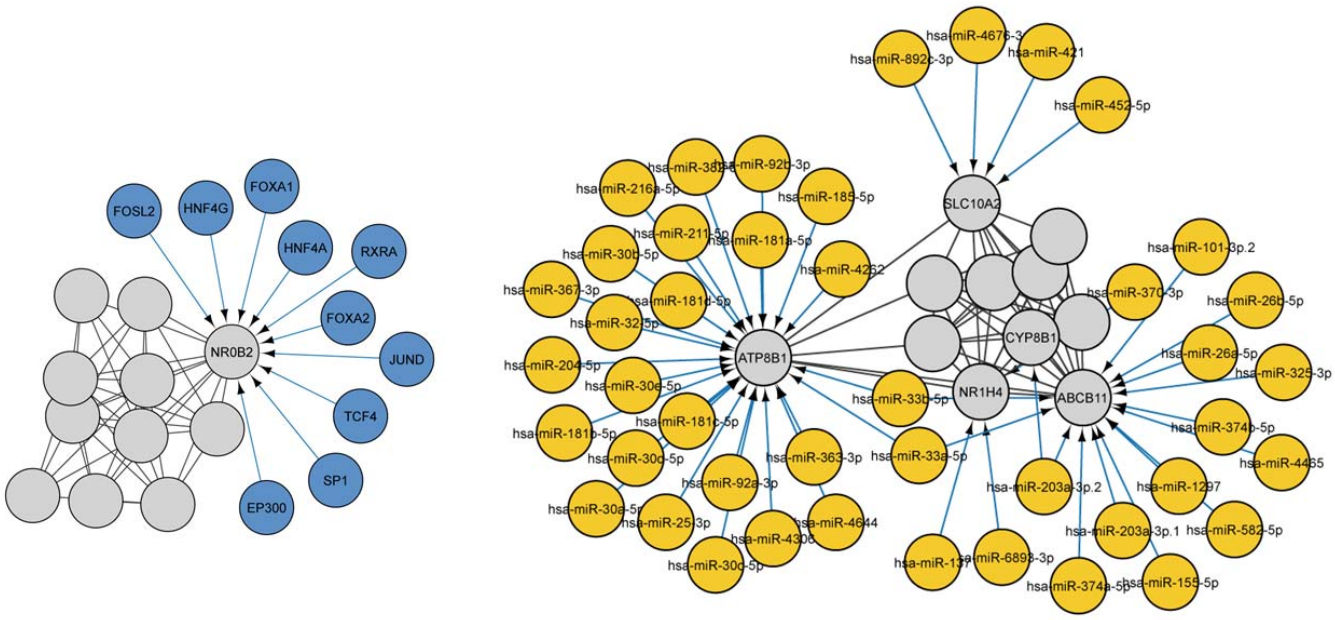


Supp-Fig-1 a,b

Protein-protein interaction network of ABCB11 gene was constructed by STRING app in Cytoscape with confidence score > 0.4 for homo sapiens. This network was constructed to analyse the relationship between ABCB11 and other proteins. Cytohubba app was used to calculate centrality of each node by MCC method. Node colour (red to yellow) represents the significance of the centrality in the group. In this analysis, we counted 11 nodes and 42 edges. These proteins majorly involved in bile acid metabolism and transport. Most of these genes are participant of more than one pathway which was expected because these pathways intersect and coregulated. We also mapped NR0B2 protein which is participate in sterol metabolism.

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Supp-Fig-2 a, b



Supp-Fig-2 a, b

Cytoscape app CyTargetlinker version 4.1.0 was used to screen the transcription factors and microRNAs using ENCODE and Target-scan databases respectively. In the screen of transcription factors of ABCB11 interaction network we observed 21 nodes and 52 edges. Among these transcription factors, FOXA have been suggested an important factor in bile duct development and lipid accumulation. HNF4A in the regulation dyslipidaemia and terminal liver failure and JUND in fibrosis development. Others can be investigated in future studies. We counted 55 nodes and 89 edges in the search of microRNA targeting the ABCB11 network. Four genes (ABCB11, ATP8B1, SLC10A2 and NR1H4) targeted by multiple microRNAs also some microRNA such as has-miR-203a-3p.2 and has-miR-203a-3p.2 target more than one gene. By nature, a microRNA can regulate several pathways therefore it would be interesting to study in future the dysregulation of these microRNAs and interaction with Identified transcription factors.

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Tables:

Table-1a Upregulated genes

Rank	Gene	UniportKB/Swiss-Prot Function
1	CXCL 10	Pro-inflammatory cytokine
2	IFIH1	Cytoplasmic sensor of viral nucleic acids
3	IFT1	IFN-induced antiviral protein
4	IFT2	IFN-induced antiviral protein
5	OASL	Antiviral activity
6	RSAD2	Induces type 1 and type 2 interferon
7	GBP2	Hydrolyzes GTP TO GMP
8	SAMD9L	Growth factor signaling
9	ISG20	Interferon -induced antiviral activity
10	CMPK2	Participate in DUTP and dCTP synthesis in mitochondria

Table-1b Downregulated genes

Rank	Name	UniportKB/Swiss-Prot Function
1	EGFR	Convert extracellular cues into appropriate cellular responses
2	PPARA	Ligand -activated transcription factor
3	CXCL12	Chemoattractant active on T-lymphocytes and monocytes
4	ENPP1	Nucleotide pyrophosphatase that genetrates diphosphate(ppi)
5	DGAT2	Triacylglycerol synthesis
6	PPAP2B	Lipid phosphatase activity
7	LPIN1	Fatty acid metabolism
8	MGLL	Converts mono-acylglycerides to free fatty acids and glycerol
9	SYT1	Calcium sensor
10	MIA3	Vesicle mediated transport

We observed that the top ranked hub genes in PPI network which were upregulated were associated with immune activity while those downregulated are associated with cell signaling and fatty acid metabolism. EGFR came first in the ranking which is a critical receptor in several cell signaling pathways.

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Table-2 Sub-network analysis

Sub-network	Seed-gene	UniportKB/Swiss-Prot Function	Diseases involved
1	CMPK2	This gene encodes one of the enzymes in the nucleotide synthesis salvage pathway that may participate in terminal differentiation of monocytic cells May participate in dUTP and dCTP synthesis in mitochondria	Retinitis Pigmentosa 39 and Thiamine-Responsive Megaloblastic Anemia Syndrome
2	ACTG1	Actins are highly conserved proteins that are involved in various types of cell motility	Deafness, Autosomal Dominant 20 and Baraitser-Winter Syndrome 2
3	SSTR2	Receptor for somatostatin-14 and -28	Neuroendocrine Tumor and Growth Hormone Secreting Pituitary Adenoma Cirrhotic liver and HCC express SSTRs.

Sub-network analysis was performed using the Molecular Complex Detection (MCODE) app from Cytoscape and CMPK2, ACTG1 and SSTR2 emerged as seed nodes among upregulated